



VU university medical center 

## Design and development of radiopharmaceuticals

Bert Windhorst  
radiopharmaceutical chemist

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### Steps in PET tracer development

- Selection of lead compounds
- Synthesis of target compounds
- Optimization of pharmacokinetic properties
- Radiolabeling with suitable radionuclide
- Binding characteristics on cells and membranes or autoradiography
- Small animal: ex vivo biodistribution, (*in vitro*, *ex vivo*) autoradiography
- Distribution studies in mice/rats with preclinical PET imaging
- Metabolism studies (N=12)
- Toxicological safety assessment. (microdosing concept)
- Set up GMP production
- Validation of PET tracer in human volunteers, including dosimetry studies, Typically N=6-12
- Proof of concept: 10 patients vs 10 controls
- Suited for clinical applications

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Complicated stuff, best to discuss with a case:  
development of [<sup>18</sup>F]PK209, a NMDA PET tracer

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## Introduction

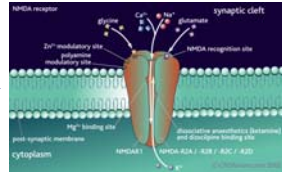


Many nervous system disorders are associated with N-methyl-D-aspartate receptor (NMDAR)

Subtype of the glutamate receptor family, contributes to many processes.

- Physiological (development, learning and memory)
- Pathological:
  - Neurodegenerative disorders, eg Alzheimer's, Huntington
  - Psychiatric disorders, eg Schizophrenia

Kotermanski, J. *Physiol* 2009, 4589-4603  
Kew, *Psychopharmacology* 2005, 179, 4-29  
Bordji, *Rev Neurosciences* 2011, 22, 285-294  
Furukawa, *Nature* 2005, 438, 185-192  
Kalia, *Lancet Neurology* 2008, 7, 742-755



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## Aim



### *In vivo* imaging of the NMDA receptor using PET

At the start of the program no suitable PET tracer available

New compounds to be designed from literature leads synthesized and evaluated *in vitro*  
high affinity compounds will be radiolabelled with  $^{18}\text{F}$  or  $^{11}\text{C}$   
evaluation of labelled compounds *in vivo* in animals

Meanwhile: [ $^{18}\text{F}$ ]GE179, published last March in *J Nucl Med*.  
Good brain uptake, homogenous distribution in the brain as can be expected.  
However not conclusive yet about the specificity of the PET signal.

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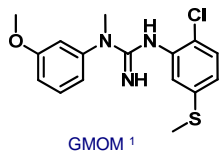
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## Lead compound



- $K_i$ : 5.2 nM ([ $^3\text{H}$ ]MK-801)<sup>1</sup>
- LogP: 2.86<sup>1</sup>

<sup>1</sup> Waterhouse *et al.* *Nucl Med & Biol* (2004) 939 and Dumont *et al.* *Bioorg. Med Chem Lett* (2002) 1883

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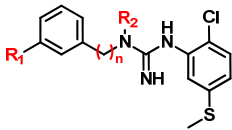
## Strategy



Guanidine structure of GMOM as template used for a new set of ion channel blockers

Possible modifications:

- introduction of 1 or 2 methylene group(s) (n)
- introduction of fluoro-alkoxy chains (R<sub>1</sub>)
- hydrogen or methyl group on guanidine (R<sub>2</sub>)



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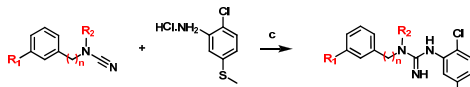
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## Chemistry (step 2)



c: chlorobenzene, 165 °C, 6h

R <sub>1</sub> =	n =	R <sub>2</sub> =
H	0, 1, 2	H, Me
O-Me	0, 1, 2	H, Me
OH	0	Me
O-CHF <sub>2</sub>	0	Me
O-CF <sub>3</sub>	0	Me

Total of  
15 compounds

Yields: 33 - 75%

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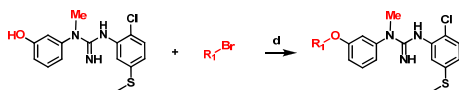
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## Chemistry (fluoro-alkanes)



d: Br(CH<sub>2</sub>)<sub>n</sub>-F, potassium carbonate, potassium iodide, DMF

R <sub>1</sub> =
CH <sub>2</sub> F
CH <sub>2</sub> CH <sub>2</sub> F
CH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> F

additional  
3 compounds

Yield: 61 - 73%

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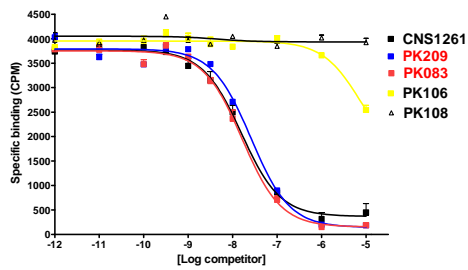
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## In vitro binding assay

Rat brain frontal cortex homogenates, with [<sup>3</sup>H]MK801



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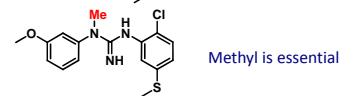
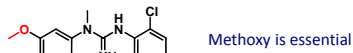
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## In vitro affinities



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## In vitro affinities

Influence of fluorine substitution in methoxy moiety

$R_1 = \text{OCH}_3$	<chem>COc1ccc(cc1)N(C)C(=O)Nc2ccc(Cl)cc2S</chem> 22 nM
$R_1 = \text{OCF}_3$	<chem>FC(F)(F)Oc1ccc(cc1)N(C)C(=O)Nc2ccc(Cl)cc2S</chem> 12 nM
$R_1 = \text{OCHF}_2$	<chem>FC(F)Oc1ccc(cc1)N(C)C(=O)Nc2ccc(Cl)cc2S</chem> 10 nM
$R_1 = \text{OCH}_2\text{F}$	<chem>FOc1ccc(cc1)N(C)C(=O)Nc2ccc(Cl)cc2S</chem> 18 nM



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## In vitro affinities



Fluoroalkanes, elongation not allowed

R1 = OCH <sub>2</sub> F		18 nM
R1 = O(CH <sub>2</sub> ) <sub>2</sub> F		155 nM
R1 = O(CH <sub>2</sub> ) <sub>3</sub> F		179 nM

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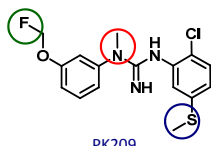
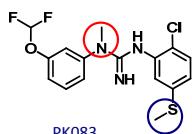
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## Selected compounds



Options for radiolabeling:

N-Methyl (carbon-11)

S-Methyl (carbon-11)

Fluoromethoxy (fluor-18)

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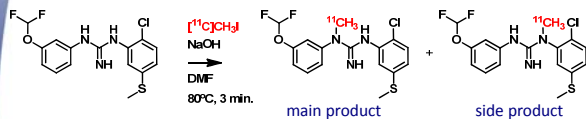
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## Radiolabelling [<sup>11</sup>C]PK083



Base	μmol	Solvent (300 μL)	RCY (HPLC)	RCY (Formulated)	Radiochemical purity
NaOH, 5M	25	DMF	51 ± 2%	9 - 26%	> 97.5 %

n = 7, decay corrected

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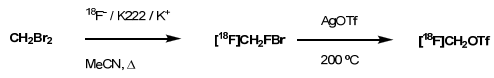
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## Radiolabelling [<sup>18</sup>F]PK209



Base	μmol	Solvent (300 μL)	RCY (HPLC)	RCY (Formulated)	Radiochemical purity
NaH	50	DMF	ND	8 - 25%	> 96 %

n = 6, decay corrected

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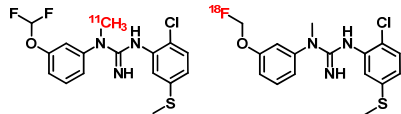
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## Specific activity and LogD<sub>7.4</sub>



Specific activity	>65 GBq/μmol	>57 GBq/μmol
LogD <sub>7.4,oct</sub>	1.76 ± 0.01	1.45 ± 0.02

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## Metabolite analysis – Methods



- Healthy male Wistar rats were injected with 90-100 MBq of tracer
- Blood samples were obtained and brains were taken out at 15 and 45 minutes post injection
- Blood plasma was passed over SPE cartridge; non-polar fraction was then eluted with MeOH and analysed by HPLC (incl. offline detection)
- Brains were homogenised in H<sub>2</sub>O, centrifuged and the supernatant was passed over a SPE cartridge; non-polar fraction was then eluted with MeOH and analysed by HPLC (incl. offline detection)

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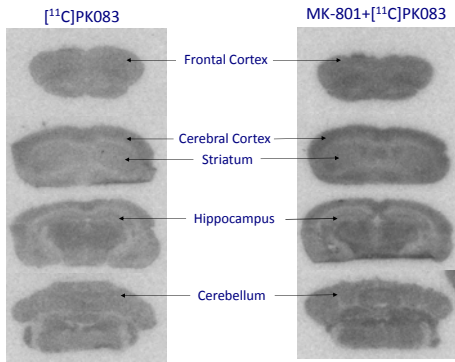
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### Ex vivo autoradiography (rats)



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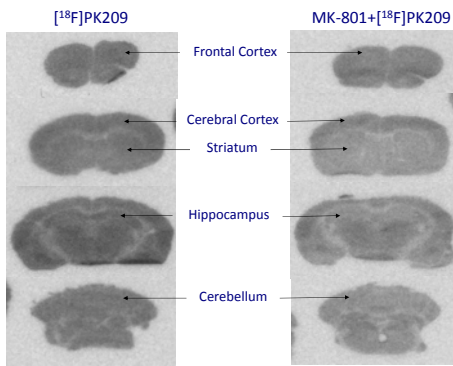
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### Ex vivo autoradiography (rats)



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### Pharmacological selectivity profile

- 10  $\mu$ M PK209 showed no significant binding to 70 of 79 targets tested
- 50-fold selectivity for NMDA ion-channel over the other 8 targets

	Target	IC <sub>50</sub> (M)	Ki (M)	nH
1	Adrenergic alpha 1A	6.2·10 <sup>-6</sup>	3.1·10 <sup>-6</sup>	1.1
2	Muscarinic M1	5.3·10 <sup>-6</sup>	4.6·10 <sup>-6</sup>	1.0
3	Muscarinic M2	4.2·10 <sup>-6</sup>	2.9·10 <sup>-6</sup>	0.8
4	Opioid $\kappa$ (KOP)	7.5·10 <sup>-6</sup>	5.0·10 <sup>-6</sup>	1.0
5	Opioid $\mu$ (MOP)	2.8·10 <sup>-6</sup>	1.2·10 <sup>-6</sup>	0.8
6	NMDA (PCP site)	4.0·10 <sup>-8</sup>	2.2·10 <sup>-8</sup>	1.5
7	sigma (1 and 2)	2.2·10 <sup>-6</sup>	1.8·10 <sup>-6</sup>	0.9
8	Ca <sup>2+</sup> channel (L, verapamil site)	4.1·10 <sup>-6</sup>	2.0·10 <sup>-6</sup>	0.7
9	Na <sup>+</sup> channel (site 2)	5.2·10 <sup>-6</sup>	4.6·10 <sup>-6</sup>	1.0

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**[<sup>18</sup>F]PK209 PET study in rhesus monkey** 

Subjects: 3 male rhesus monkeys, aged 17-19 years, weight: 7.6-13.5 kg

Sedation with midazolam (0.3 mg/kg) + medetomidine (60 µg/kg), anaesthetised with propofol (0.2 mg/kg/min)

T1-weighted structural MRI scans in 3T Siemens Trio scanner

120 min dynamic PET scans in the ECAT HRRT scanner

Continuous and discrete manual blood samples from the femoral artery

Blocking scans performed 1-2 weeks after baseline. NMDAs blocked with MK-801 (0.3 mg/kg), administered 30 min before radiotracer injection

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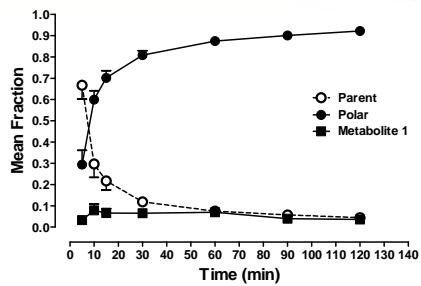
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**Plasma metabolism in rhesus monkey** 



No effect of MK-801 on metabolism, no defluorination  
<sup>[<sup>18</sup>F]PK209 shows fast metabolism in plasma, comparable to rats.</sup>

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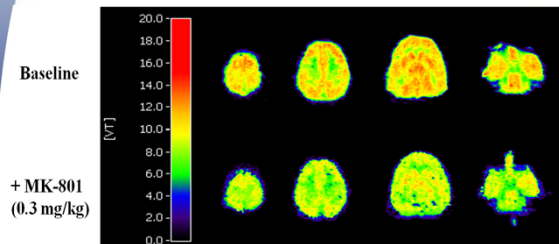
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**Logan V<sub>T</sub> parametric images, subject 1** 



Images presented through the transaxial plane  
 Smoothed by isotropic Gaussian filter of 2 mm at full-width, half maximum

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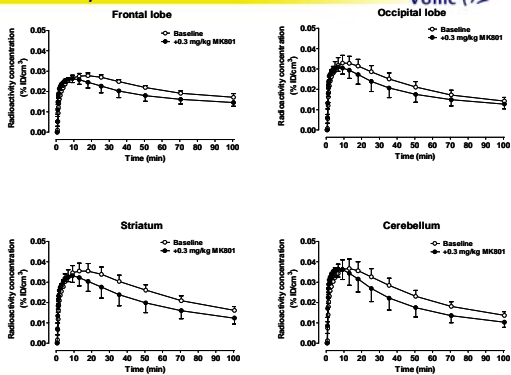
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## Time-activity curves



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## Microdosing toxicity



Performed at Advinus, India according to EMEA/CPMP/SWP/2599/02/Rev 1, June 2004 and EMEA/CPMP/ICH/286/95 June 2009

Wistar rats, 1 and 14 day toxicity at a dose of 95 µg/kg (1000 times the expected tracerdose)

5 males, 5 females per time point, vehicle control groups

clinical signs, mortality, changes in body weight and food intake, terminal hematology, coagulation and clinical chemistry, organ weights, gross pathology and histopathological examination of liver and lungs.

Results: no toxicity observed.

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## Conclusions



2 compounds were selected for radiolabeling  
From a series of NMDA noncompetitive ion channel blockers,

Reliable radiosynthesis was established

Ex vivo autoradiography showed unfavorable results with [<sup>11</sup>C]PK083

[<sup>18</sup>F]PK209 PET in rhesus monkey showed modest reduction of the signal after MK-801 administration.

PK209 was shown to be safe in a microdosing toxicity study

Further characterisation in humans is warranted

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## Acknowledgements



Chemistry: Pieter Klein & Hans  
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PI: Bart van Berckel  
Supervision: Ronald Boellaard &  
Adriaan Lammertsma

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